# Acremonium exuviarum sp. nov., a lizard-associated fungus with affinity to Emericellopsis

Lynne Sigler<sup>1\*</sup>, Alga Zuccaro<sup>2</sup>, Richard C. Summerbell<sup>3</sup>, Julian Mitchell<sup>4</sup> and Jean A. Paré<sup>5</sup>

<sup>1</sup>University of Alberta Microfungus Collection and Herbarium, Edmonton, Alberta, Canada T6G 2E1; <sup>2</sup>Institut für Mikrobiologie, Technische Universität Braunschweig, Spielmannstr. 7, D-38106, Braunschweig, Germany; <sup>3</sup>Centraalbureau voor Schimmelcultures, Fungal Biodiversity Centre, KNAW, Uppsalalaan 8, 3584 CT Utrecht, The Netherlands; <sup>4</sup>School of Biological Sciences, University of Portsmouth, King Henry Building, King Henry I Street, Portsmouth P01 2DY, U.K.; <sup>5</sup>Department of Surgical Sciences, School of Veterinary Medicine, University of Wisconsin, 2015 Linden Drive West, Madison, WI 53706, U.S.A.

\*Correspondence: lynne.sigler@ualberta.ca

**Abstract:** In a survey of cycloheximide-tolerant fungi growing from shed reptile skins, an *Acremonium*-like fungus was isolated that was distinctive in producing relatively large conidia in chains from phialides that tended to collapse after forming a number of conidia. Phylogenetic analysis based on ribosomal internal transcribed spacer and  $\beta$ -tubulin sequences revealed that the isolate represented an undescribed and relatively phylogenetically isolated member of the clade containing the teleomorph genus *Emericellopsis* as well as related anamorphs in *Acremonium* and *Stanjemonium*. The species is here described as *Acremonium exuviarum*. It has been isolated only on a single occasion and its ecology is unknown. Discovery of such new species in the pharmaceutically important *Emericellopsis* clade is potentially of practical significance.

Taxonomic novelty: Acremonium exuviarum Sigler, Zuccaro, Summerbell & Paré sp. nov.

Key words: Acremonium, Emericellopsis, Corucia zebrata, skink, veterinary

## INTRODUCTION

An Acremonium-like fungus was isolated from shed dorsal skin of a captive female Solomon Island, or prehensile-tailed, skink (Corucia zebrata Gray) during a survey of the mycobiota from healthy reptiles (Paré et al. 2003). The objective of the survey was to examine freshly shed exuviae from 127 healthy captive reptiles for the presence of the Chrysosporium Corda anamorph of Nannizziopsis vriesii (Apinis) Currah, recently identified as an agent of cutaneous mycosis in reptiles (Paré et al. 1997, Nichols et al. 1999, Thomas et al. 2002). Although that survey used a medium containing cycloheximide to selectively recover Chrysosporium species and onygenalean fungi, there was a high frequency of recovery of saprotrophic fungi. Acremonium species, for example, were recovered at a frequency of 13 % from shed skin samples. The Acremonium isolate from the skink demonstrated unusual features, so its identification was investigated in some detail. The relatively large round conidia and branching phialide structure suggested a possible Emericellopsis F.H. Beyma lineage and the isolate was included in a broader study of terrestrial- and marine-associated Emericellopsis species and related anamorphs (Zuccaro et al. 2004, this volume). Morphological and phylogenetic analyses confirm that the skink isolate is distinct and that it has affinities within Emericellopsis.

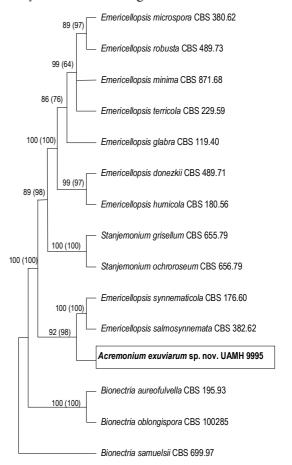
# **MATERIALS AND METHODS**

Details on the methods for sampling of shed skin samples, specimen submission and culture are provided in Paré et al. (2003). The Solomon Island skink was an adult wild-caught female (coded CZ-F) housed at the Steinhart Aquarium, San Francisco, CA. Shedding skin was removed with forceps aseptically from the dorsal area and the right flank. The isolate described here was obtained from the dorsal skin and deposited as UAMH 9995 in the University of Alberta Microfungus Collection and Herbarium (UAMH), Edmonton, Alberta, Canada. A subculture is deposited in Centraalbureau voor Schimmelcultures, Utrecht, the Netherlands, as CBS 113360. Colony descriptions are based on potato-dextrose agar (PDA, Difco Laboratories, Detroit, MI, U.S.A.) at 25 and 30 °C, 2 % malt extract agar (containing Difco malt extract) at 22 °C, and oatmeal-salts agar (OAT; Kane et al. 1997). Herbarium specimens were prepared by growing the fungus on PDA and OAT overlaid with a cellophane membrane (Carmichael 1961). Colour names are according to Kornerup & Wanscher (1978). Cycloheximide tolerance was evaluated on Mycosel agar (MYC, Difco). Microscopic morphology was examined from Riddell slide culture preparations incubated at 25 °C and mounted in lacto-fuchsin or polyvinyl alcohol, following the method and recipes described in Kane et al. (1997). The nutrient medium was cereal agar made with Heinz (H.J. Heinz, Pittsburgh, PA, U.S.A.) mixed cereal, also manufactured as Pablum mixed cereal.

DNA extraction, amplification and sequencing methods are as described in Zuccaro et al. (2004). Sequences of the nuclear ribosomal internal transcribed spacer (ITS) region (555 characters), inclusive of the intervening 5.8S rDNA, as well as a portion of the β-tubulin locus (347 characters) were aligned manually (GENEDOC; Nicholas et al. 1997) and combined to form a matrix of 902 characters. The final alignment comprised fifteen taxa including three Bionectria Speg. species used as outgroup. Relationships among strains included in the present phylogenetic analysis are shown in Fig. 1. These are a subset of those examined by Zuccaro et al. (2004; provenance provided in their Table 1). Each genetic region was partitioned within the matrix to allow independent analyses and partition homogeneity tests (Farris et al. 1994, Swofford 2000). All phylogenetic analyses were performed using PAUP 4.0b10 (Swofford 2000). Maximum parsimony settings included: heuristic searches with random sequence addition (10-50 replicates) using the tree-bisection reconnection (TBS) branch-swapping algorithm with and without successive weighting (Farris 1969). Bootstrap analyses and partition heterogeneity tests were performed using up to 1000 replicates. Maximum likelihood (ML) with quartet puzzling (Strimmer & von Haeseler 1996) was performed to account for any rate heterogeneity existing between the partitions. Substitution rate-matrix parameters and nucleotide frequencies were estimated from the data and from site-specific rates calculated according to the general time-reversible + site-specific rates (GTR + SS) substitution model implemented in PAUP. Likelihood calculations were made using the least squares method and the number of puzzling steps varied between 10 000 and 50 000.

#### RESULTS

Acremonium exuviarum sequences displayed a high degree of homology with those of *Emericellopsis* and Stanjemonium W. Gams, O'Donnell, Schroers & C.M. Chr. species (Fig. 1). ITS region similarities ranged from 95-99 % within the group, with the sequence from A. exuviarum matching others at the 96-97 % level. The values of similarity dropped to 76–77 % when this sequence was compared to those from species of Bionectria, the sister group of Emericellopsis (Rossman et al. 2001). The molecular link to Emericellopsis is more pronounced when the  $\beta$ tubulin "intron 3" sequences are compared. Emericellopsis and Stanjemonium species have a very short intron 3 sequence, varying in length between 49 and 52 base pairs. The intron 3 sequence from A. exuviarum is also very short and has a high similarity to that of *E. synnematicola* P.N. Mathur & Thirum. and *E. salmosynnemata* Grosklags & Swift.



**Fig 1.** The relationship of *Acremonium exuviarum* sp. nov with *Emericellopsis* based upon combined nuclear ribosomal internal transcribed spacer (ITS)-5.8S and  $\beta$ -tubulin sequence analysis. Strict consensus cladogram of two most parsimonious trees of 351 steps. Bootstrap values are shown above the branches with quartet puzzling supports given in brackets. GenBank accession numbers for sequences are AY882947 ( $\beta$ -tubulin) and AY882946 (ITS).

Separate maximum parsimony analyses of the ITS and β-tubulin genes generated similar topological trees, but did not resolve the placement of some species (data not shown). Partition homogeneity tests between ITS1 and ITS2, and between the whole ITS region and the β-tubulin gene, revealed no significant differences (P = 0.3 and P = 0.37 respectively) between the distributions of tree lengths, allowing these datasets to be combined. Maximum parsimony analysis of the combined dataset generated two most parsimonious trees of length 351 steps (confidence index [CI] = 0.821, retention index [RI] = 0.867, rescaled consistency index [RC] = 0.711 and homoplasy index [HI] = 0.179; parsimony-informative characters [PIC] = 180) from a single tree-island. Figure 1 shows the strict consensus of these two trees and the bootstrap support values for each node. A consensus tree of similar topology was generated after ML-quartet puzzling analysis. This analysis estimated the relative rates for the two genetic regions (0.733 for the ITS and 1.4 for the  $\beta$ -tubulin gene) so that any rate heterogeneity between them could be taken into account. In both analyses, *A. exuviarum* formed a strongly supported clade with *E. synnematicola* P.N. Mathur & Thirum. and *E. salmosynnemata* Grosklags & Swift (92 % bootstrap support and 98 % quartet puzzling support).

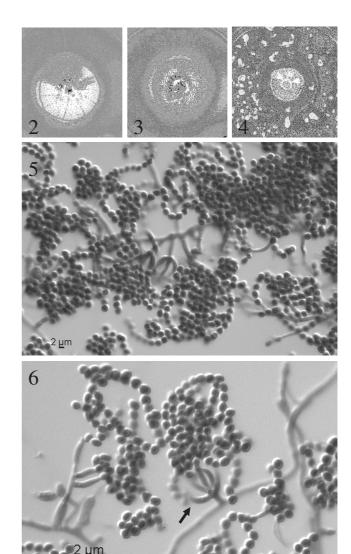
*Acremonium exuviarum* Sigler, Zuccaro, Summerbell & Paré, **sp. nov.** MycoBank MB500101. Figs 2–8.

Etymology: exuvia (L) referring to cast off skins.

Coloniae in agaro moderatim rapide crescunt 25–30 °C, cremeae vel pallide aurantiacae, sulcatae et elevatae, vel planae et zonatae, saepe partes texturae variae formantes, pars aversa pallide grisea; incrementum tardum ad 37 °C. Hyphae hyalinae, septatae, leves, angustae, 0.5–2 µm latae, interdum fila formantes. Phialides simplices vel (2–)3(–4) verticillatae in conidiophoro brevi non distincto; tenuiter curvatae, 3.5–20(–40) × 1.3–3 µm; cellulae terminales postea collabuntur. Conidia catenis vel capitulis mucidis connexa, hyalina, levia, ovoidea vel subglobosa, 3–4 × 2–3 µm. Chlamydosporae absunt; teleomorphosis ignota.

*Holotypus*: Colonia exsiccata (UAMH 9995); cultura viva (UAMH 9995; CBS 113360).

Colonies on PDA after 10 d at 25 and 30 °C similar, 3.2-3.5 cm diam, yellowish white (4A2) to orangewhite (6B2), slightly elevated at the center, with few to many radial folds, felty, finely zonate, sometimes forming sectors of white funiculose hyphae or glabrous, reverse pale grey, lacking diffusible pigments. Growth slower at 22 °C (2.8 cm diam on PDA; 2 cm on 2 % MEA). Colonies on 2 % MEA thin, hyaline. On OAT, colonies after 10 d at 25°C 2.5 cm diam, flat, zonate, powdery, heavily sporulating, pale orange (5A3) in the centre, becoming thin, orange white (5A2) at the margin; colonies at 22 °C thinner, not zonate. Growth strongly inhibited at 35 °C (5 mm after 14 d on PDA). Good growth on medium with cycloheximide. Hyphae in slide culture preparations (as per Kane et al. 1997) are hyaline, septate, smooth, narrow, 0.5-2 µm wide, sometimes aggregating in strands. Conidiogenous cells are phialides attached singly on the mycelium or borne in verticils of 2-4 (commonly 3) on short undifferentiated conidiophores. Phialides are cylindrical, slightly tapered at the tip, without visible collarettes, often slightly curved especially when borne in verticils, 3.5–20(–40)  $\mu m$  long (av. 9.2  $\mu m$ ) and 1.3–3  $\mu m$  wide (av. 1.7 um); terminal cells sometimes collapsing.



**Figs 2–4.** Colonies of *Acremonium exuviarum* (UAMH 9995) after 10 d. 2. PDA 25 °C. 3. PDA 30 °C. 4. OAT 25 °C. 5-6. Conidia formed in chains. Note phialides are single or borne in verticils (arrow) of (2-)3(-4) on short undifferentiated conidiophores. Preparations are stained with lactofuchsin. Scale bar = 2  $\mu$ m.

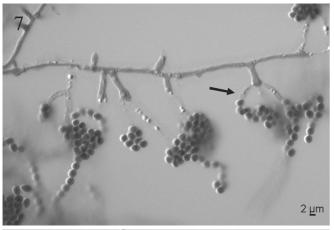
*Conidia* are formed in loose chains which may collapse into heads and are hyaline, smooth, ovoid to subglobose,  $3-4\times 2-3$  µm. *Chlamydospores* and teleomorph not observed.

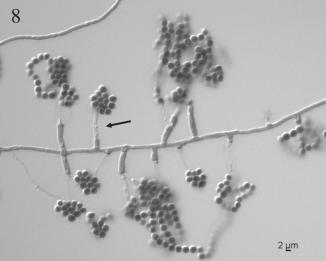
## **DISCUSSION**

The morphological features of *A. exuviarum* are unusual among members of the *Emericellopsis* clade. Conidia are formed in loose chains and phialides appear to collapse in age. Collapsing conidiogenous cells are described for *Stanjemonium* species, but these cells produce a single conidium and collapse soon after this conidium is mature. This may be an adaptation to limited availability of free moisture in the desert or semi-desert soils from which most *Stanjemonium* isolates have been collected. Consideration was given to placing our new taxon in *Stanjemonium*,

but this was rejected on both morphological (multiconidial vs. uniconidial phialides) and phylogenetic grounds. The *Stanjemonium* species grouped in a subclade well separated from the branch bearing *A. exuviarum*. Based on a broad analysis of *Emericellopsis* and related anamorphs, Zuccaro *et al.* (2004) have argued that *Stanjemonium* could either be taken up for all named anamorphs in the *Emericellopsis* clade, or it could be one of four genera accommodating anamorphs within the clade. We follow their decision to retain *Acremonium* for most anamorph species with affinities to *Emericellopsis*. The occurrence of collapsing phialides in *A. exuviarum*, however, provides support for their observation that these may not be distinctive at the generic level.

Acremonium exuviarum is the only member of the Emericellopsis clade known to produce conidia in chains. The production of catenulate conidia by a single member of this clade support the suggestion by Gams (1971) that this feature has little or no taxonomic significance above the species level in the hypocrealean species that comprise the bulk of the genus Acremonium.





Figs 7, 8. Acremonium exuviarum (UAMH 9995) in slide culture preparation stained with lacto-fuchsin showing disintegration of terminal conidiogenous cells. Scale bar =  $2 \mu m$ .

This parallels the situation seen in related hypocrealean genera such as *Fusarium* Link (O'Donnell *et al.* 1998), *Lecanicillium* Zare & W. Gams (2001) and *Clonostachys* Corda (Schroers 2001), where very closely related taxa may form conidia either in chains or in mucoid heads.

Acremonium exuviarum is represented only by its type, so little is known about its ecology. *Emericellop*sis species are traditionally considered to be soil inhabitants; however recent work has indicated that these organisms are associated with a range of substrates, including brown algae (Zuccaro et al. 2003, 2004) and wood (Lai Ka Pang, pers. comm.). As shown by Zuccaro et al. (2004), the terrestrial species of Emericellopsis tested so far express cellulases and polyphenoloxidase, suggesting a potential to be involved in plant and wood decay. Although the expression of cellulase was not tested in the same manner for A. exuviarum, this fungus was slightly cellulolytic when grown on OAT overlaid with a cellophane membrane (Carmichael 1961). Acremonium exuviarum was isolated from an adult wild-caught female skink in active shed. It is unknown how long the animal had been in captivity at the Steinhart Aquarium in San Francisco. Veterinarians and keepers were requested to sample only from animals that appeared healthy and free of skin lesions and to collect the samples aseptically. The animal may have been exposed to the fungus on material provided for climbing; this often consists of pieces of wood. Corucia zebrata, a member of the Scincidae, is native to the tropical rain forests of the Solomon Islands where it is arboreal. Large powerful claws and a long prehensile tail enable it to cling to trees and it is often found in trees of old growth forests (de Vosjoli 1993). However, the possibility that A. exuviarum simply derived from air spora settled on the lizard's skin cannot be excluded.

The *Emericellopsis* clade is of unusual pharmaceutical significance. It includes significant sources of cephalosporin C and related compounds (Backus & Orpurt 1961) as well as of other metabolites of interest such as bergofungin peptaibols (Berg *et al.* 1999). It is hoped that elucidation of new biodiversity within this group will lead to investigation of the taxa involved for compounds of potential interest in medicine and industry.

# **ACKNOWLEDGEMENTS**

We thank Mieke Starink-Willemse and Arien van Iperen for technical assistance in the cultivation and sequencing of *A. exuviarum*, and Mr. Michael Hertwig-Jaksch for assistance with the Latin. Financial support from the Natural Sciences and Engineering Research Council of Canada to L. Sigler, and from the Association of Reptilian and Amphibian Veterinarians to J. Paré is gratefully acknowledged.

### REFERENCES

- Backus MP, Orpurt PA (1961). A new *Emericellopsis* from Wisconsin, with notes on other species. *Mycologia* **53**: 64–83.
- Berg A, Schlegel B, Ihn W, Demuth H-U, Gräfe U (1999). Isolation and structural elucidation of new peptaibols, bergofungins B, C and D, from *Emericellopsis donezkii* HKI 0059A. *Journal of Antibiotics* **52**: 666–669.
- Carmichael JW (1961). Dried mold colonies on cellophane. *Mycologia* **55**: 283–288.
- De Vosjoli P (1993). The general care and maintenance of the prehensile-tailed skinks. The herpetological library series. Advanced Vivariums Systems, Inc., Lakeside, U.S.A.
- Farris JS (1969). A successive approximations approach to character weighting. *Systematic Zoology* **18**: 374–385.
- Farris JS, Källersjö M, Kluge AG, Butt C (1994). Testing significance of incongruence. *Cladistics* **10**: 315–319.
- Gams W (1971). *Cephalosporium-artige schimmelpilze* (*Hyphomycetes*). Gustav Fischer Verlag, Stuttgart, Germany.
- Gams W, O'Donnell K, Schroers H-J, Christensen M (1998). Generic classification of some more hyphomycetes with solitary conidia borne on phialides. *Canadian Journal of Botany* **76**: 1570–1583.
- Kane J, Summerbell RC, Sigler L, Krajden S, Land G (1997) Laboratory handbook of dermatophytes. A clinical guide and laboratory manual of dermatophytes and other filamentous fungi from skin, hair and nails. Star Publishing Co., Belmont, CA.
- Kornerup A, Wanscher JH (1978). *Methuen handbook of colour*, 3<sup>rd</sup> ed. Methuen, London, U.K.
- Nicholas KB, Nicholas HB Jr, Deerfield DW II (1997). GeneDoc: Analysis and visualization of genetic variation. *Embnet News* **4**: 14. (http://www.psc.edu/biomed/genedoc/ebinet.htm)
- Nichols DK, Weyant RS, Lamirandel EW, Sigler L, Mason RT (1999). Fatal mycotic dermatitis in captive brown tree snakes (*Boiga irregularis*). *Journal of Zoo and Wildlife Medicine* **30**: 111–118.
- O'Donnell K, Cigelnik E, Nirenberg HI (1998). Molecular systematics and phylogeography of the *Gibberella fuji-koroi* species complex. *Mycologia* **90**: 465–493.

- Paré JA, Sigler L, Rypien KL, Gibas CF. (2003). Survey for the *Chrysosporium* anamorph of *Nannizziopsis vriesii* on the skin of healthy captive squamate reptiles and notes on their cutaneous fungal mycobiota. *Journal of Herpetological Medicine and Surgery* **13**:10–15.
- Paré JA, Sigler L, Hunter DB, Summerbell RC, Smith DA, Machin KL (1997). Cutaneous mycoses in chameleons caused by the *Chrysosporium* anamorph of *Nannizziopsis vriesii* (Apinis) Currah. *Journal of Zoo and Wildlife Medicine* **28**: 443–453.
- Rossman AY, McKemy JM, Pardo-Schultheiss RA, Schroers H-J (2001). Molecular studies of the *Bionectriaceae* using large subunit rDNA sequences. *Mycologia* **93**: 100–110.
- Schroers, H-J (2001). A monograph of *Bionectria* (*Ascomycota, Hypocreales, Bionectriaceae*) and its *Clonostachys* anamorphs. *Studies in Mycology* **46**: 1–214.
- Sigler L, Flis A (1998). Catalogue of the University of Alberta Microfungus Collection and Herbarium, 3<sup>rd</sup> ed. University of Alberta, Edmonton, Canada [available online at www.devonian.ualberta.ca/uamh].
- Swofford D. (2000). *PAUP\**. *Phylogenetic analysis using parsimony* (\* *and other methods*). Sinauer Associates, Sutherland, Massachusetts, Version 4.
- Strimmer K, von Haeseler A. (1996). Quartet puzzling: a quartet maximum likelihood method for reconstructing tree topologies. *Molecular Biology and Evolution* **13**: 964–969.
- Thomas AD, Sigler L, Peucker S, Norton JH, Nielan A. (2002). *Chrysosporium* anamorph of *Nannizziopsis vriesii* associated with fatal cutaneous mycoses in the saltwater crocodile (*Crocodylus porosus*). *Medical Mycology* **40**: 143–151.
- Zare R, Gams W (2001). A revision of *Verticillium* section *Prostrata*. IV. The genera *Lecanicillium* and *Simplicillium* gen. nov. *Nova Hedwigia* **73**: 1–50.
- Zuccaro A, Schulz B, Mitchell JI (2003). Molecular detection of ascomycetes associated with *Fucus serratus*. *Mycological Research* **107**: 1451–1466.
- Zuccaro A, Summerbell R, Gams W, Schroers H-J, Mitchell JI (2004). A new *Acremonium* species associated with *Fucus* spp., and its affinity with a phylogenetically distinct marine *Emericellopsis* clade. *Studies in Mycology* **50**: 283–297.

SIGLER ETAL.